

Removal of Amoxicillin From Wastewater Onto Activated Carbon: Optimization of Analytical Parameters by Response Surface Methodology

Dose-Response:
An International Journal
July-September 2024:1–15
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DOI: 10.1177/15593258241271655
journals.sagepub.com/home/dos



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Abstract

Antibiotics are widely used in veterinary and human medicine, but these compounds, when released into the aquatic environment, present potential risks to living organisms. In the present study, the activated carbon (AC) used for their removals is characterized by FT-IR spectroscopy, BET analysis and Scanning Electron Microscopy (SEM) to determine the physicochemical characteristics. Response surface methodology (RSM) and Box-Behnken statistical design (BBD) were used to optimize important parameters including pH (2–12), temperature (20–45°C), and AC dose (0.05–0.20 g). The experimental data were analyzed by analysis of variance (ANOVA) and fitted to second-order polynomial using multiple regression analysis. The optimal conditions for maximum elimination of Amoxicillin (Amox) are (Dose: 0.124 g, pH 5.03 and 45°C) by applying the desirability function (df). A confirmation experiment was carried out to evaluate the accuracy of the optimization model and maximum removal efficiency ($R = 89.999\%$) was obtained under the optimized conditions. Several error analysis equations were used to measure goodness of fit. Pareto analysis suggests the importance of the relative order of factors: pH > Temperature > AC dose in optimized situations. The equilibrium adsorption data of Amox on Activated Carbon were analyzed by Freundlich, Elovich, Temkin and Langmuir models. The latter gave the best correlation with q_{\max} capacities of 142.85 mg/g ($R^2 = 0.999$) at 25°C is removed from solution. The adsorption process is dominated by chemisorption and the kinetic model obeys a pseudo-second order model ($R^2 = 0.999$).

Keywords

amoxicillin, adsorption, activated carbon, optimization, isotherm, modeling

Introduction

Although pharmaceutical compounds such as antibiotics are necessary for animals and humans, their overuse has become a global problem due to the resistance of pathogens to these drugs.¹ The release of pharmaceuticals into natural waters is one of the main environmental concerns; these compounds mainly come from the pharmaceutical industry and hospitals, and threaten water resources.² Even in small quantities, antibiotics and emerging contaminants generate microbial resistance³ and municipal wastewater treatment plants are not efficient enough to eliminate them completely.⁴ To this end, adsorption remains attractive and showed its effectiveness in removing these pollutants from wastewater, as it is simple to

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Received 23 April 2024; accepted 26 June 2024

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Data Availability Statement included at the end of the article



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apply with low cost, it is currently well known that (Amox) is used in the modern medicine, possessing significant ecotoxicity. Amoxicillin is a broad-spectrum beta-lactam antibiotic that belongs to penicillin class used as veterinary medicine for treatment of bacterial infections encountered in gastrointestinal and systemic infections. Relatively few reports have been published on their elimination. Amoxicillin (Amox) is a widely consumed antibiotic; its structure is based on β -lactam ring, responsible for its high bacterial resistance against microorganisms.^{5,6} To assess the risks associated with medical wastewater, it is currently well known that amoxicillin possesses significant ecotoxicity.⁷ It is a broad-spectrum beta-lactam belonging to the penicillin class used in veterinary medicine for the treatment of bacterial and gastrointestinal and systemic infections.⁸ It is commonly prescribed for infections caused by bacteria.⁹ Recent studies have reported the presence of amoxicillin at concentrations between ng/L and mg/L in domestic¹⁰ and industrial waters¹¹ and many antibiotics have a toxic nature towards algae and other organisms which may have a long-term indirect effect on ecological sustainability.¹² Indeed, due to its bioaccumulation in natural environments, Amox can induce toxic effects influencing the fauna and the flora, which contributes to the destruction of the ecosystem.¹³ It is included on the World Health Organization's list of essential medicines, as one of the most effective medicines in a health system.¹⁴ The presence of antibiotics in the environment has been associated with chronic toxicity and their resistance to antibiotics in bacterial species.¹⁵ Therefore, its elimination by an effective process is crucial before its release into the aquatic environment. To date, many methods have been tried to eliminate antibiotics in conventional treatment plants, like filtration,^{16,17} photocatalytic degradation,¹⁸⁻²⁰ ozonation,²¹ chlorination,²² biological degradation,²³ and advanced oxidation processes (AOPs)^{24,25} have been applied to remove drugs from sewage streams. However, the adsorption remains attractive and Activated carbon production requires carbon precursors which are usually agricultural wastes, such as rice husk, peach pits, date palm, lotus stem, rice straw, almond shell, macadamia nut shell, walnut shells, tea waste, sugar beet pulp, peanut shells, wood chips, and olive stones etc... In this study, commercial activated carbon is used as an adsorbent for the removal of Amox in aqueous solution due to its porosity. Experimental design for variable optimization has a few advantages i) it reduces the number of experiments, resulting in the use of less material and reagents and ii) provides information about interactions between variables.²⁶ The Response surface methodology (RSM) with BBD design is a widely used mathematical optimization the analytical parameters (temperature, adsorbent dose and pH) with the desirability method (df) through to simultaneous analysis of interactions of several parameters with the minimum of experimental trials. BBD is a spherical type design which consists of central point and middle points of the edges of the cube circumscribed on the sphere,^{27,28} the mutual influence of the parameters is studied by the method Pareto and

the analysis of variance (ANOVA) method is used as a criterion for the validation of the mathematical model. Response surface methodology (RSM) with BBD design is a mathematical optimization of analytical parameters (temperature, adsorbent dose and pH of the medium) with the desirability method (df) through the simultaneous analysis of the interactions of several parameters with the minimum of experiments.

Materials and methods

Reagents

The antibiotic studied is amoxicillin (high purity, Sigma-Aldrich), the characteristics and developed formula of which are grouped into Table 1, it was purchased from in analytical purity. Different concentrations of solutions were prepared by diluting a stock solution in distilled water. The adsorbent used for its elimination is a commercial Granular activated carbon (AC) supplied by Dinâmica Química Contemporânea (Diadema, Brazil). Before use, the adsorbent was thoroughly washed with water to remove carbon powder and surface impurities, followed by drying at 100°C for 48 h. The size fraction between 2.00 and 2.38 mm was selected for all adsorption tests.

Characterization of the Activated Carbon

The FT-IR spectroscopy, performed using a Bruker spectrometer (Perkin Elmer 200P), was used to obtain the qualitative identification of the functional groups present on the adsorbent surface; the spectra were obtained between 4000 and 400 cm^{-1} (resolution of 4 cm^{-1}).²⁹ The N_2 adsorption/desorption isotherms were performed using a Tri Star II 3020 Micrometrics instrument, obtain the textural characterization of the CCP.600 and CCP.700 adsorbent. The adsorbent samples were firstly dried (12 h) at 120°C in an internal oven of the apparatus, and then the surface analyzer was run at -196°C for the N_2 isotherms.³⁰ The pore size distribution was calculated by the DFT method using the N_2 adsorption branch.³¹ The surface areas were calculated using the BET method.³² The textural surface of AC was captured by Scanning electron micrograph (SEM), taken with different resolutions thanks to FEI Quanta650FEG equipment.³³ The Point of Zero Charge (pHpzc) was determined by pH titration procedures.³⁴ 50 cm^3 of KNO_3 (0.01 M) solutions was poured into erlenmeyer flasks; the pH of solution within each flask was adjusted between 2 and 9 by addition of HCl or NaOH (0.1 M) solution. Then, 0.15 g of adsorbent was added to the flasks and the final pH was measured after 48 h. The pHpzc is defined as the intersection point where the curve pH_{final} vs $\text{pH}_{\text{initial}}$ crosses the line $\text{pH}_{\text{final}} = \text{pH}_{\text{initial}}$.

Table 1. Technical Characteristics of the Studied Amoxicillin (Amox).

Product Specification		Structural Formula
Product name	Amoxicillin	
Product number	A8523	
CAS number	26787-78-0	
Storage temperature	Cooler/Refrigerated	
Molecular formula	C ₁₆ H ₁₉ N ₃ O ₅ S	
Formula weight	365.40	
Appearance (form)	Crystalline powder	
Water (by Karl Fischer)	11.5-14.5 %	
Assay (anhydrous basis)	95.30-102.0 %	

Adsorption Experiments

The batch equilibration method was followed for the optimization of the Amox adsorption according to the CCD matrix. For this a precise amount of adsorbent was mixed with 0.1 L of (Amox) solution. The mixture was stirred with a magnetic agitation at 150 rpm at fixed contact time (40 min). Then, the separation was carried out by centrifugation at 6000 rpm during 10 min and the (Amox) concentrations retained in the biosorbent phase were analyzed by UV-Vis spectrophotometry. The experiments were carried out under variable conditions: pH, initial Amox concentration, AC adsorbent and temperature. HCl and NaOH (1.0 mol/L) were used to adjust the solution pH. To eliminate any interference with the adsorbates, blank tests were performed alongside the biosorption experiments by replacing the (Amox) solutions with water.

Adsorption Process Optimization by Response Surface Method

In the present study, a central composite design (3²) was used to evaluate the influence toward the Amox adsorption onto AC. The BBD statistical experimental design and RSM were used to study the effects of the four independent variables on the response function. RSM is a specific, combined and mathematical tool for designing the experiment, modeling the construction process and monitoring the effects and impacts of variables and operating conditions. It is useful for optimization to examine the optimal conditions to take into account the response to lower or higher end results of a particular process.^{35,36} To study their impact on Amox removal, the most significant variables of adsorption were considered: pH (X₁), temperature (X₂), and adsorbent dose (X₃). The low, center and high levels of each variable are denoted by -1, 0 and 1, respectively (Table 2). A total of 30 experiments were designed using the RSM method and optimized by a central composite design. Each experimental run was analyzed twice to avoid the effects of external factors, and the average values were considered. The percent removal of Amox for each run was reported within ± 0.5 % precision, the following

Table 2. Independent Variables and Their Levels Used for Box-Behnken Design.

Variable, Unit	Factors	Levels		
		-1	0	1
pH solution	X ₁	2	7	12
Temperature T (°C)	X ₃	25	40	45
Adsorbent dose D (g/L)	X ₂	0.0225	0.1112	0.2

polynomial response second-order equation was derived to explain the system behavior:

$$Y = a_0 + \sum_{i=1}^K a_i x_i + \sum_{i=1}^k \sum_{j=1}^k a_{ij} x_i x_j + \sum_{i=1}^k a_{ii} x_i^2 \quad (1)$$

where Y is the predicted response which represents the removal efficiency R(%) of (Amox) adsorbed, a₀ a constant coefficient, x_i and x_j variables. a_i, a_{ii} and a_{ij} are the interaction coefficients for linear, quadratic and second order terms, respectively. The removal efficiency by AC was calculated as follows:

$$R(\%) = \frac{(C_0 - C_e) \cdot 100}{C_0} \quad (2)$$

C_e is the initial and equilibrium concentration of the Amox (mg/L) and C₀ the initial concentration. The experimental matrix of CCD was proposed by Minitab Software 16 which was also used to determine the regression analysis of designed model and graphical analysis of each response. The total number of experiments calculated using the following equation:

$$N = k^2 + k + Cp \quad (3)$$

(N = 30) is the total number of runs, k (=3) the number of independent variables, Cp (=3) the number of experiment repeated at center points and α (= 0.0) the distance of axial point from center. ANOVA was used to analyze the statistical significance of each variable and the adequacy of the resulting regression equation. The experimental design matrix found

according to the CCD as well as the answer are given in Table 3.

Modeling and Statistical Analysis

In order to verify the quality of the mathematical model through the experiments, we used the Student tests (T value), and Fisher tests (F value) as well as the coefficient of determination (R^2), obtained by the Minitab 16 software.

The Student's test makes it possible to statistically verify the hypothesis of equality of the expectation of two random variables following a normal distribution and an unknown variance. It is also used to test the nullity of a coefficient in the context of a linear regression. The Fischer test is a statistical hypothesis which verifies the equality of two variances, by the ratio of the two variances which does not exceed a certain theoretical value, sought in the Fisher table. In our case, the software gives the ratio between the adjustment variance and that of the experimental error.

The R^2 coefficient is an indicator for judging the quality of a linear, simple or multiple regressions. For a value between 0 and 1, it measures the adequacy between the model and the observed data.

ANOVA was performed to verify the statistical significance of the experimental design and establish interactions between independent variables and response effects.³⁷ It subdivides the total variation into two components, with variation associated with either the model or experimental error.³⁸ F - and P -values (prob > F , confidence interval = 95 %) for the model were obtained. ANOVA is a statistical approach that subdivides the total change of the data set into ingredient parts concomitant with a defined source to test the target hypotheses on the model parameters.³⁹ The value of Fisher's F test, which is the ratio between the mean square of the model and the residual error, allows this comparison and was applied in this study.⁴⁰

Adsorption Equilibrium Isotherms

Adsorption isotherms are of primary importance for optimizing the removal of hazardous molecules. The equilibrium relationship between adsorbent and adsorbate is best explained, and the maximum uptake capacity is obtained from the adsorption isotherm. To assess the AC performance, different equations and isotherms exist, out of which the Langmuir,⁴¹ Freundlich,⁴² Temkin⁴³ and Elovich⁴⁴ isotherms are presented in the following equations respectively:

$$q_e = \frac{q_{\max} \cdot K_L \cdot C_e}{1 + K_L \cdot C_e} \quad (4a)$$

$$R_L = \frac{1}{1 + K_L \cdot C_0} \quad (4b)$$

$$q_e = B_T \cdot \ln(A_T \cdot C_e) \quad (5a)$$

Table 3. Box-Behnken Experimental Design Matrix and Responses (R_{exp} (%) and R_{pred} (%) for Elimination of (Amox) Onto AC.

Run	X_1	X_2	X_3	R_{exp} (%)	R_{pred} (%)
1	2	0.1125	45	82.5	85.62
2	7	0.1125	35	85.0	85.58
3	12	0.1125	25	26.6	23.57
4	2	0.0250	35	75.0	72.61
5	7	0.0200	25	54.4	60.12
6	12	0.1125	45	58.6	60.43
7	7	0.2000	45	72.2	67.97
8	2	0.2000	35	60.18	61.28
9	12	0.0250	35	26.82	25.72
10	7	0.1125	35	85.25	85.42
11	7	0.0250	45	87.35	86.62
12	12	0.0200	35	45.2	47.59
13	7	0.1125	35	86.0	85.42
14	7	0.0250	25	26.72	30.95
15	2	0.1125	25	60.8	58.97

R_{exp} (%) is determined experimentally.

R_{pred} (%) is calculated according to the polynomial formula.

$$B_T = R \cdot T \cdot q_{\max} / \Delta E_T \quad (5b)$$

$$q_e = K_F \cdot C_e^{1/n} \quad (6)$$

$$\frac{q_e}{q_{\max}} = K_E \cdot C_e \cdot \exp\left(-\frac{q_e}{q_{\max}}\right) \quad (7)$$

q_{\max} (mg/g) is the monolayer adsorption capacity and K_L the Langmuir separation constant related to the free adsorption energy (L/mg), A_T , B_T , ΔE_T (kJ/mol), K_F , n and K_E (L/mg) are the Temkin, Freundlich and Elovich constants. Furthermore, the isothermal models were applied under optimal parameter conditions. The Langmuir model is the best known and widely applied, it is represented in the non linear form.

Results and Discussions

Surface Functional Groups of the Activated Carbon

The functional groups of AC contribute significantly to its adsorption capacity, with selectivity to attract cations to the adsorbent surface.⁴⁵ These functional groups are consisted mainly of acidic and basic groups which affect the surface charges and adsorption properties of activated carbon. Adsorption on activated carbon therefore not only relies upon its pore structure as the change of surface charges is also a crucial factor affecting the adsorption capacity. The FTIR spectrum can reveal the functional groups on activated carbon surface qualitatively based on the characteristic energy absorbed for each bond in certain groups,⁴⁶ although some particular peaks may overlap. The FTIR spectrum gives the functional groups on the activated carbon surface based on the energy absorbed for each bond,⁴⁶ although some particular peaks may overlap. The

presence of C = O peaks around 1685 cm^{-1} comes from the carboxylic functional groups, as well as the chromene and pyrone structures (Figure 1). The peak 1025 cm^{-1} comes from C–O bonding while the broad band ($3550\text{--}3200\text{ cm}^{-1}$) is assigned to be O–H functional groups for carboxyl and phenol. The presence of phenol is confirmed by the presence of peak around 1578 and 1610 cm^{-1} which was the absorption energy of benzene rings.

Physical Characteristics of Adsorbent

The isotherms of AC (Figure 2) reveals different structures for activated carbon, the curves is rather confused and shows rapid N_2 adsorption at low relative pressure ($P/P_0 < 0.4$), indicating that the activated carbon is mainly made up of micro pores (pore size $< 20\text{ \AA}$).⁴⁷ The BET surface area of activated carbon determined on the basis of nitrogen sorption is $1092.951\text{ m}^2/\text{g}$.

Analyses of the AC Surface Morphology

To confirm the adsorption of Amoxicillin, SEM images were captured before and after adsorption (Figure 3) where we notice a difference in the arrangement of the cavities. The SEM micrographs of the active carbon before and after adsorption are shown in; AC presents a micro-porous structure with different pore diameters with a rough surface and numerous protrusions. After adsorption, the AC surface became smoother, with the roughness significantly reduced and the pores less visible, indicating an adsorption on both the surface and inside the pores. The SEM images also reveal that the exterior surfaces of AC are filled with more or less homogeneous cavities of different sizes and shapes. The cavities are the external pores and represent the main channels of AC to access the internal surface (micropores and mesopores). The highest magnification clearly shows that the adsorbent surface contains a considerable number of pores with a high probability that Amox molecules are adsorbed inside these pores.

Point of Zero Charge (pH_{pzc})

pH_{pzc} is a point where an adsorbent has a zero charge on its surface and the presence of H^+ or OH^- ions in solutions may change the surface charges of the adsorbent. Above pH_{pzc} , the surface functional groups on adsorbents are protonated by the excess H^+ ions; on the contrary, below pH_{pzc} , the surface functional groups are deprotonated by the OH^- ions presence in the solution.⁴⁸ pH_{pzc} of our activated carbon is equal to 5.00.

Mechanism of Action of Amoxicillin

Amox is commonly used due to the broad spectra in terms of its mechanism of action as it stops the proliferation of different

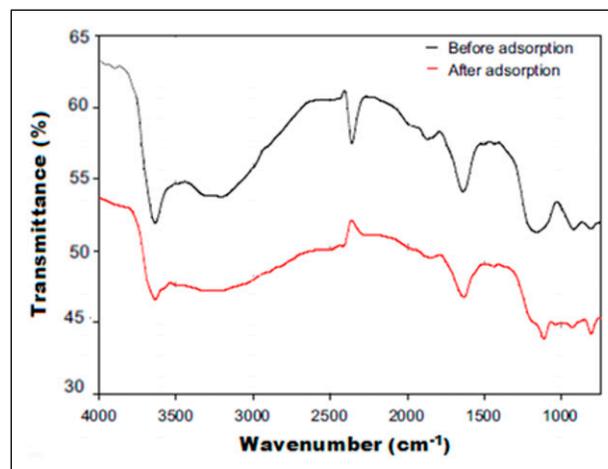


Figure 1. FTIR spectra for activated carbon before and after Adsorption.

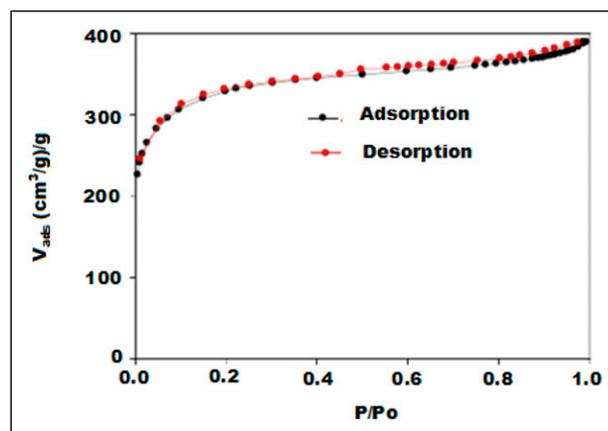


Figure 2. Nitrogen sorption and desorption isotherms for activated carbon.

bacteria.^{49,50} In general, the penicillin inhibits a bacterial, an enzyme involved in the synthesis of the bacterial cell wall.⁵¹ The β -lactam rings are involved in the inhibition mechanism. Penicillin covalently binds to the active site of the enzyme, leading to an irreversible inhibition. It is worth while to mention that Amox has amphoteric properties,⁵² due to its three main functional groups (Figure 4), $-\text{COOH}$ ($\text{pK}_{\text{a}1} = 2.7$), $-\text{NH}_2$ ($\text{pK}_{\text{a}2} = 7.4$) and $-\text{OH}$ ($\text{pK}_{\text{a}3} = 9.6$).^{53,54} The mechanisms of interaction Amox/AC should occur according to the proposals reported in the literature,^{55,56} it is compatible with the polar groups that exist in the AC surface and Amox (Figure 5).

- i) π - π interactions (π bonds of AC with π bonds of antibiotic)
- ii) Donor-acceptor complex formation between the surface carbonyl groups (electron donors) and the aromatic ring of Amox acting as the acceptor.

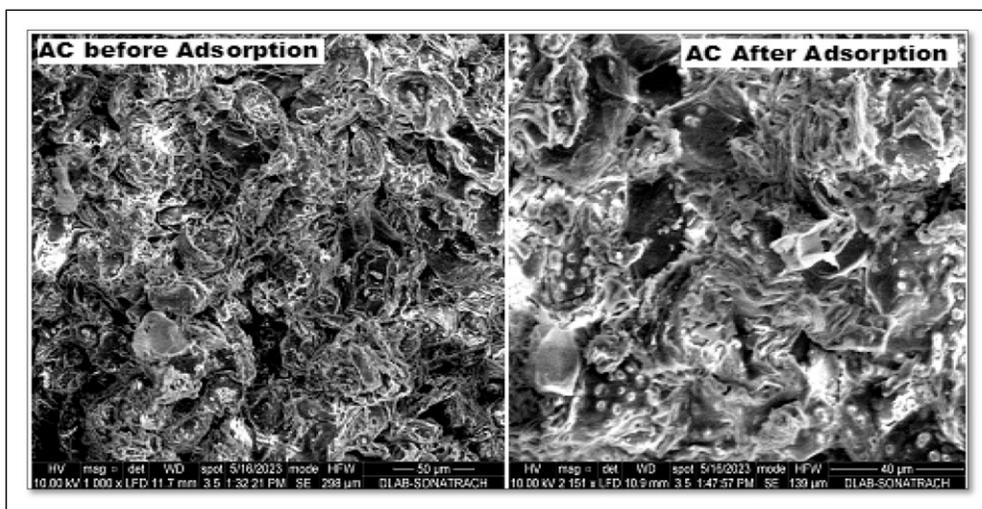


Figure 3. Microscopic images of the Activated Carbon before and after adsorption.

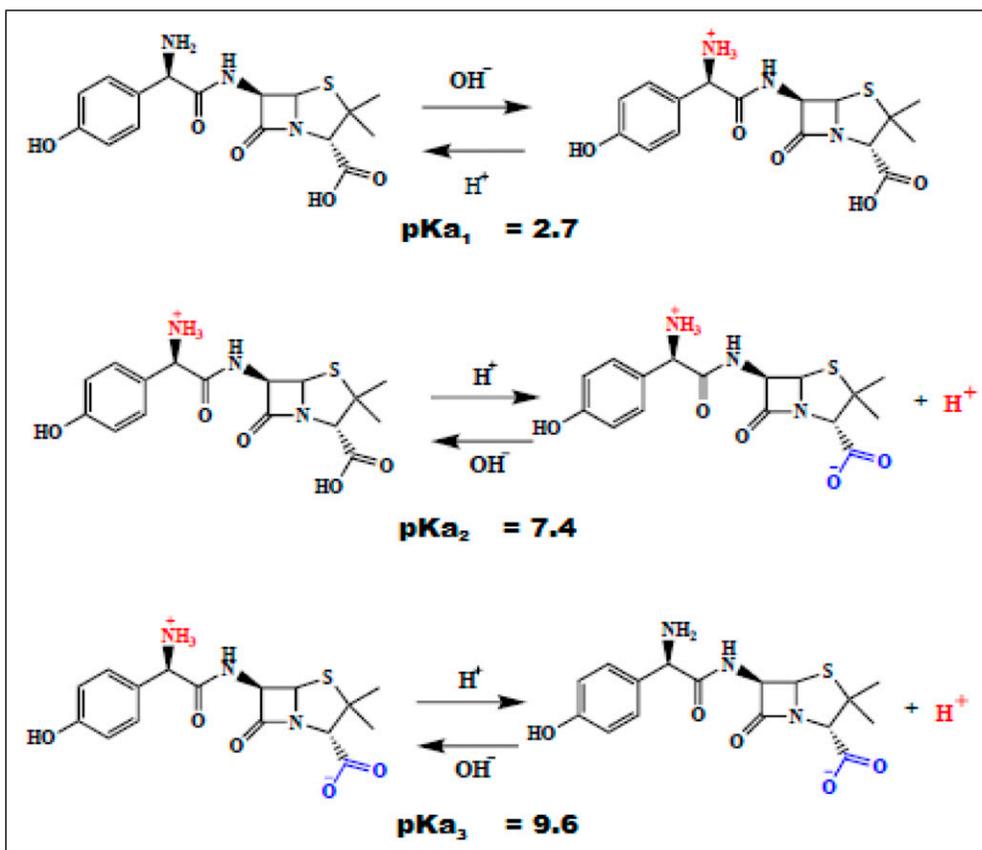


Figure 4. Mechanism of action of amoxicillin.

- iii) Hydrogen bonds between the -OH and -NH groups of Amox and the -OH and -NH groups of AC

At pH_{pzc}, AC gave the highest removal performance. At pH_{pzc} of AC (4.91), Amox exists in its neutral form

(the pH range where amoxicillin exists in neutral form is 3-6)⁵⁷ Similar observations were also noted by other researchers⁵⁸ At pH_{pzc} of adsorbent ($\text{pKa}_1 < \text{pH}_{\text{sol}} < \text{pKa}_2, \text{pKa}_3$), amine functional groups of Amox present in the form of $-\text{NH}_3^+$ and carboxyl functions can be found in the form of $-\text{COO}^-$. Therefore, the electrostatic force

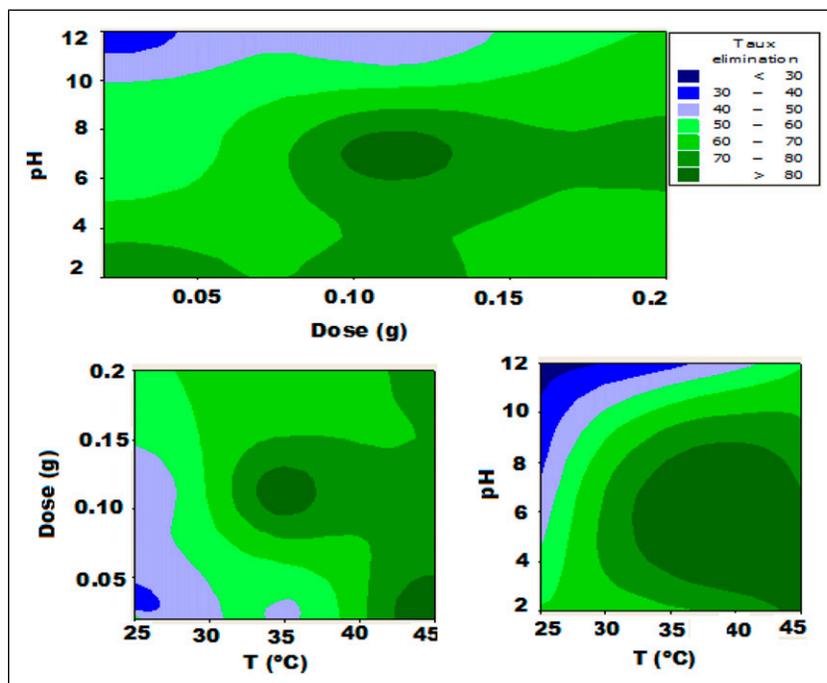


Figure 5. The contours plots of (solution pH - adsorbent dosage), (adsorbent dosage - Temperature) and (solution pH - Temperature).

would enhance the adsorption since AC contains both positive and negatively functional groups which attract the Amox groups. Another possible mechanism, also worth to be mentioned is the reaction between the amine functions of Amox with carbonyl functional groups on activated carbon. C = O in carbonyl groups are converted by the NH₂ groups to C = N while H₂O is released in solution.

Experimental Design Analysis by Central Composite Design

The effect of temperature, adsorbent and pH on Amox uptake by AC was investigated using RSM. Fifteen sets of experiments with different experimental conditions were generated by RSM. The relationship between the percentage uptake of Amox and the three significant factors were illustrated by a second-order polynomial:

$$\begin{aligned}
 R(\%) = & 85.1501 - 13.6013 \text{ pH} - 0.2308 \text{ Dose} \\
 & + 14.3376 \text{ T} - 18.9188 \text{ pH}^2 - 11.1191 \text{ Dose}^2 \\
 & - 9.3063 \text{ T}^2 + 6.4555 \text{ pH} \cdot \text{Dose} + 2.5750 \text{ pH} \cdot \text{T} \\
 & - 8.0680 \text{ Dose} \cdot \text{T}
 \end{aligned}$$

According to the model, the Temperature independent variable, and the interaction pH*Dose and pH*T have a positive sign, indicating a good impact on the response, with enhanced elimination of Amox. Validation of this model was carried out using the desirability function (df) where the most desirable experimental conditions generated by the software were achieved. The results

obtained for the comparison of experimental yields and those calculated by the mathematical model during the eliminations of Amox on activated carbon are shown in Figure 6 and Table 3 to confirm the correlation of the results, a plot of the $R_{\text{exp}} = f(R_{\text{pred}})$ curve is carried out (Figure 7), the latter clearly shows that the experimental results and those deduced by the model are in perfect correlation ($R^2 = 0.984$). The CCD responses were analyzed and the results of analysis of variance, and RSM were analyzed by Minitab software (version 16) for adsorption study. To determine the comparative significance (Figure 8) of each term in the model, Pareto analysis was applied using the following equation:

$$P_i = \left(\frac{a_i^2}{\sum a_i^2} \right) \cdot 100 \quad (8)$$

Were a_i is the regression coefficient of individual process variable. The influence of factors and their interactions is studied through the postulated model which indicates which coefficients should be retained and which could be eliminated from the final mathematical model. To evaluate the importance of a coefficient, statistical theory compares this coefficient to its standard deviation thus indicating the T -Student which determines the P -value.

An ANOVA analysis (Table 4) was generated from the experimental results according to conditions designed by RSM. To determine whether the effect of a coefficient is statistically significant, we compare the P -value with the alpha confidence level (= 0.05): if P -value < 0.05, the effect is significant; above 0.05) the effect is insignificant.

The significance of the effects of the factors can also be identified by the Pareto analysis, in which the effects are

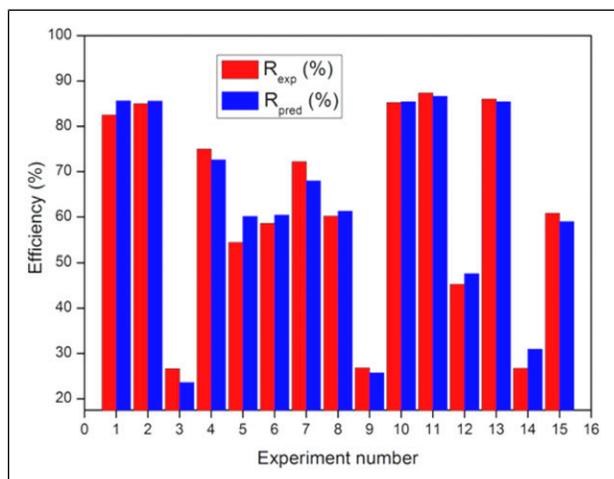


Figure 6. Comparison of elimination yields.

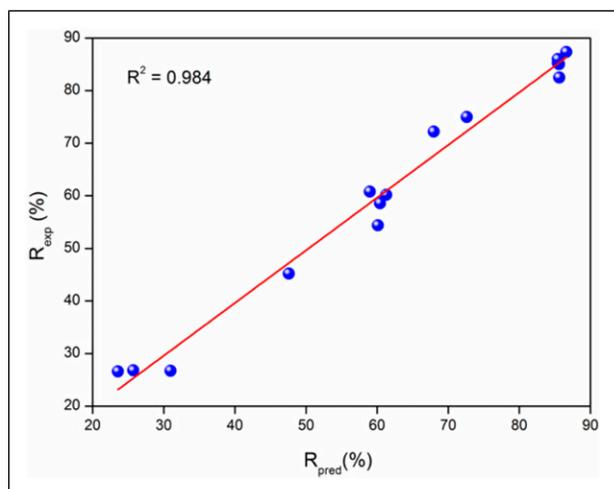


Figure 7. Plot of the experimental and predicted response.

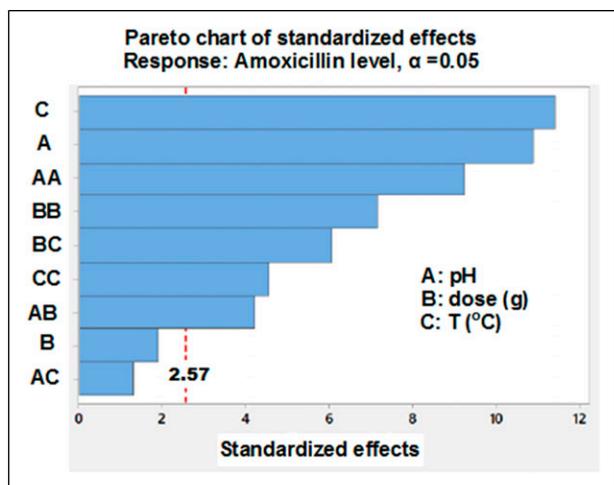


Figure 8. Pareto graphic analysis.

ordered from most influential to least. The reference line for statistical significance is automatically indicated by the software. The large T -value and small P -value indicates a high significance of regression models. Some interactions for Amox pollutant (pH, T , and pH*pH), the P -value of less than 0.05 indicates that the experimental factors are significant at 95 % confidence level. But the interaction pH*D, pH*T, D*T, T*T, D*D and D present P -value more than 0.05 indicates that the experimental factors are not significant.

The effects of factors are identified by Pareto analysis, in which they are ranked from most influential to least, the reference line for statistical significance is indicated by the software. The large T - and small P -value indicate significance of the regression models. Some interactions (pH, T and pH*pH), P -value less than 0.05 indicates these factors are significant at 95% confidence level. But the interaction pH*D, pH*T, D*T, T*T, D*D and D has a P value greater than 0.05 indicating that these experimental factors are not significant.

Main Effects Diagram for Amoxicillin

Figure 9 shows the simultaneous influence of factors on the elimination rate of Amox. The adsorption rate increases with pH with a maximum pH of 7 then decreases. Adsorption with dose increases in the range (0.025 - 0.1125 g) and then decreases by this value. Temperature also has a positive effect on the elimination of Amox.

Diagram of the interaction effects of pH and dose factors

For a dose of 0.025 g requires an acidic medium to achieve an elimination rate of 75%, while a maximum elimination rate at pH 7 and an adsorbent dose of 0.1125 g, then it decreases by increasing the dose up to 0.2 g (Figure 10).

Diagram of the Interaction Effects of the Factors pH and Temperature T ($^{\circ}C$)

For a dose of 0.025 g requires an acidic medium to achieve an elimination rate of 75%, while a maximum elimination rate at pH 7 and an adsorbent dose of 0.1125 g, then it decreases by increasing the dose up to 0.2 g; the results are summarized in Figure 11.

Optimization by RSM and with Desirability Function

The Minitab software offers a graphical route to study the interactions of the terms of the 2D contour model, adapted to binary interactions. Contour plots of the second order polynomial equation in two independent variables (pH - temperature,

Table 4. Analysis of Variance (ANOVA) for Response Surface Quadratic Model.

Source	Coef	SE Coef	T	df	Seq SS	Adj SS	Adj MS	F	P
Regression				9	6453.86	6453.86	717.10	5.79	0.034
Constant	85.1501	6.422	13.260						
Linear				3	4020.80	2569.31	856.44	6.92	0.031
pH	-13.6013	4.980	-2.731	1	1838.00	923.48	923.48	7.46	0.041
D	-0.2308	6.222	-0.037	1	195.66	0.17	0.17	0.00	0.972*
T	14.3376	4.979	2.880	1	1987.13	1026.56	1026.56	8.29	0.035
Square				3	2234.42	1653.06	551.02	4.45	0.071*
pH*pH	-18.9188	6.318	-2.995	1	1022.42	1110.03	1110.03	8.97	0.030
D*D	-11.1191	8.025	-1.386	1	862.53	237.64	237.64	1.92	0.225*
T*T	-9.3063	6.309	-1.475	1	349.47	269.37	269.37	2.18	0.200*
Interaction				3	198.65	198.65	66.22	0.53	0.678*
pH*D	6.4555	7.979	0.809	1	44.22	81.04	81.04	0.65	0.455*
pH*T	2.5750	5.563	0.463	1	26.52	26.52	26.52	0.21	0.663*
D*T	-8.0680	7.937	-1.016	1	127.90	127.90	127.90	1.03	0.356*
Residual error				5	618.94	618.94	123.79		
Lack-of-Fit				2	449.48	449.48	224.74	3.98	0.143
Pure error				3	169.45	169.45	56.48		
Total				14	7072.80				

Significance: * Parameter is not significant.

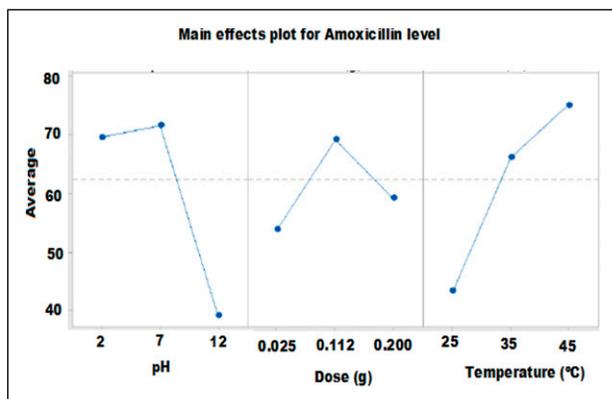


Figure 9. Main effects diagram for Amoxicillin.

pH - adsorbent dose, pH - temperature, pH - Co); the other parameters are kept at fixed values. Contour plots of the second order polynomial equation in two independent variables (pH and temperature, pH and Co, pH and adsorbent dose for Amox). It can also include the main factor and the interaction impacts of two independent factors.⁵⁹

Examination of the 2D curves of the binary variables allows us to delineate the following optimal conditions to obtain maximum elimination of Amox (pH 6, T = 45°C; and AC dose = 0.125 g/L). The desirability function (Figure 12) makes it possible to simultaneously know the optimal parameters (pH 5.03, T = 45°C; AC dose = 0.125 g/L with R (%) = 89.99) of the variables of input that determines the optimal value. Performance levels for one or more responses occur in two stages:

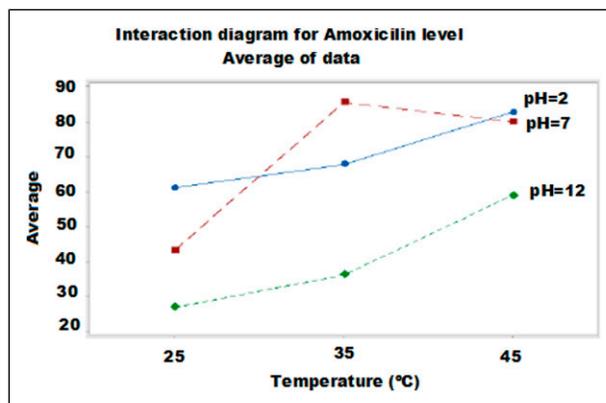


Figure 10. Diagram of the interaction effects of pH and temperature T (°C).

- i) Find the levels of the independent variables that simultaneously produce the most desirable, predicted responses on the dependent variables.
- ii) Maximize overall desirability with respect to controllable factors, the desirability function approach was initially introduced by Harrington.⁶⁰

Adsorption Isotherms

Adsorption isotherms were applied to evaluate the amount of Amox adsorbed on AC surface and the type of interaction. The graphical representation of the linearized forms of the four isotherms (Figure 13) allowed us to calculate the constants of the different models (Table 5). The evaluation of the physical

condition of the data at equilibrium, based on the coefficient R^2 as a criterion, shows the applicability of the Langmuir and Temkin models to interpret the data with a maximum adsorption capacity q_{max} ($= 142.85 \text{ mg/g}$, $R^2 = 0.999$). Langmuir's model is the best suited. Note that R_L indicates the type of isotherm: Irreversible ($R_L = 0$), Favorable ($0 < R_L < 1$), Linear ($R_L = 1$) or unfavorable ($R_L > 1$). In our case, the R_L values are less than 1, thus confirming that the adsorption is well described by the Langmuir isotherm which suggests one of the following three main hypotheses:

- i) The adsorption sites have identical adsorption energy.
- ii) The adsorbate occupies only one site on the surface, with no interaction between the adsorbate molecules
- iii) The adsorption is mostly confined to the adsorbent surface.

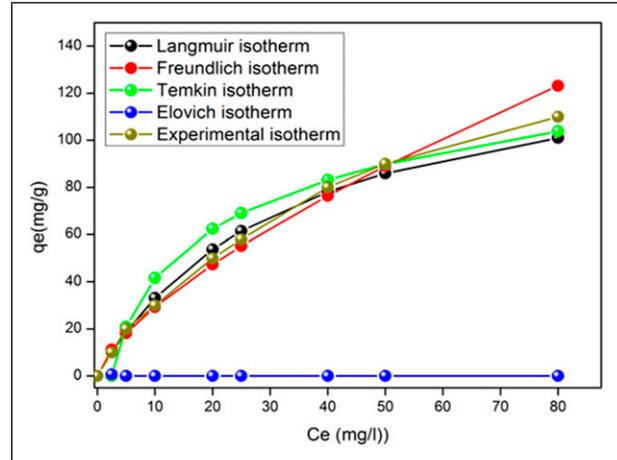


Figure 13. Isotherms Adsorption for the different models in the optimum conditions.

Comparison with Other Adsorbents Used for the Removal of Amoxicillin

It is instructive for comparative purposes to report the maximum adsorption of amoxicillin from some available

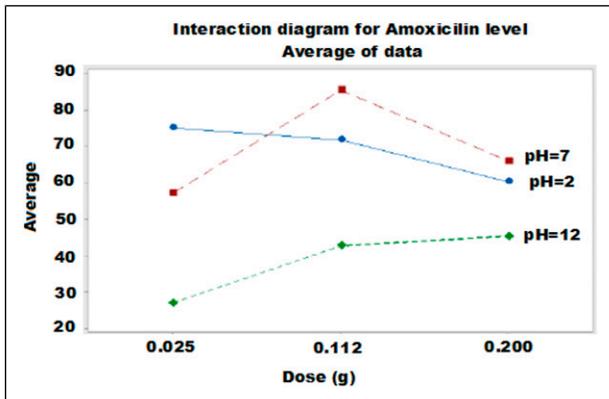


Figure 11. Diagram of the interaction effects of pH and dose factors.

Table 5. Different Parameters for Adsorption Isotherm Models for the Removal of Amox Onto AC.

Model	Parameters	Amox
Langmuir	Q_{max} (mg/g)	142.85
	K_L (L/mg)	0.0302
	R^2	0.999
	χ^2	0.009
Freundlich	K_F (mg/g) (L/mg) ^{1/n}	5.989
	1/n	0.69
	R^2	0.938
	χ^2	6.726
Temkin	A_T B_T	0.402
	E (j/mol)	29.93
	R^2	11.196
	χ^2	0.989
Elovich	Q_{max} (mg/g)	1.623
	K_E	3.239
	R^2	1.856
	χ^2	0.948
		13.522

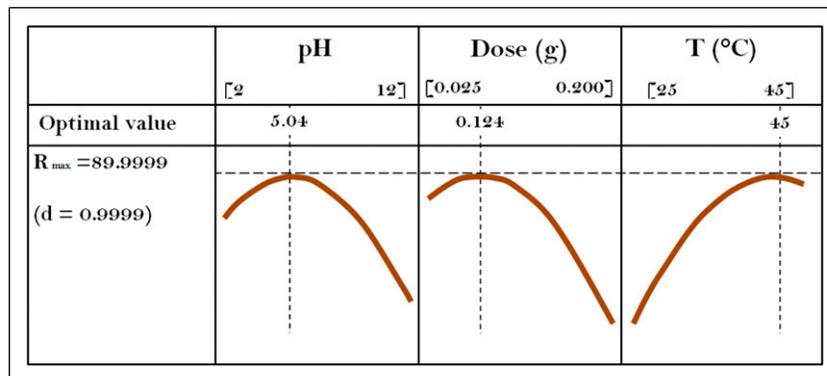


Figure 12. Predicted responses using desirability function.

Table 6. Comparison of Maximum Adsorption Capacity With Other Adsorbents Reported in the Literature.

Adsorbent	Q _{max} (mg/g)	References
Bentonite	53.93	61
Bentonite	49.99	.
Bentonite	47.37	.
Activated carbon	186.43	.
Activated carbon	221.86	.
Activated carbon	189.55	62
Capsules of cashew of Para 600°C	289.2	.
Capsules of cashew of Para 600°C	392.1	.
Capsules of cashew of Para 700°C	292.0	.
Capsules of cashew of Para 700°C	348.9	.
Zinc oxide coated carbon nanofibers	156	63
NaOH-activated carbon	571	64
Activated carbon	4.4	65
Activated carbon from <i>Arundo donax</i> Linn	345	66
Multi-walled carbon nanotubes	22.9	67
Magnetic multi-walled carbon nanotubes	50	68
Iron oxide nanoparticles supported on mesoporous MCM-41	25	69
Magnetic adsorbent prepared from olive kernel (MA-OK)	238	70
Mn-impregnated activated carbons	122-132	71
Cellulose from flax noil	183	72
Commercial activated carbon	142.85	This studies

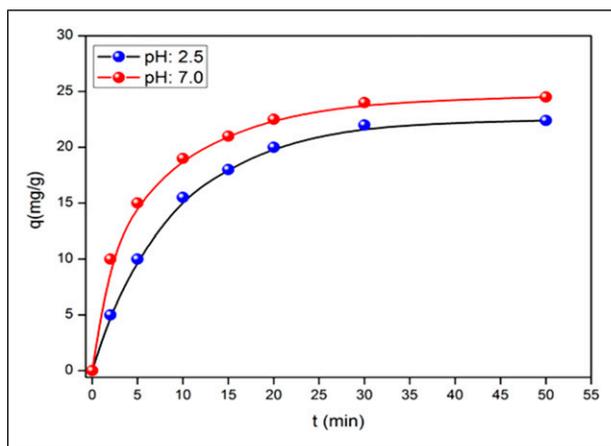


Figure 14. Effect of pH on the Adsorption Kinetic.

adsorbents (Table 6). We can see that the adsorption performance of amoxicillin on AC is ranked well compared to other research with a maximum capacity q_{max} of 146.85 mg/g, relatively interesting. The elimination of this waste has a double impact: the recovery of this waste in water treatment and the protection of the natural environment and the ecosystem.

Desorption constitutes an intermediate step towards the regeneration of the adsorbent. It is an essential tool to evaluate the reuse of any adsorbent for large-scale applications, due to ecological concerns and sustainable development needs. This study gave encouraging results. We wish to extend our study

Table 7. Different Parameters for Adsorption Kinetics Models for the Removal Of Amox Onto AC.

pH	Q _{ex} (mg/g)	Second Order	First Order
pH = 2.5	22.40	$K_2 = 8.03 \times 10^{-3} \text{ g/mg.min}$	$K_1 = 3.178 \text{ min}^{-1}$
pH = 7.0	27.027	$Q_{cal} = 26.32 \text{ mg/g}$ $R^2 = 0.995$	$Q_{cal} = 23.98 \text{ mg/g}$ $R^2 = 0.980$
		$K_2 = 10.10 \times 10^{-3} \text{ g/mg.min}$ $Q_{cal} = 27.027 \text{ mg/g}$ $R^2 = 0.995$	$K_1 = 3.007 \text{ min}^{-1}$ $Q_{cal} = 20.23 \text{ mg/g}$ $R^2 = 0.980$

to column adsorption tests under conditions applicable to the treatment of industrial effluents.

Adsorption Kinetic Modeling

The kinetics of Amox adsorption is crucial to determine the optimized operating conditions for a large-scale process. It gives the absorption rate of the adsorbate, controls the residual time of the overall process and predicts both the adsorption rate and the adsorption design. Different models have been proposed to study the behavior of adsorbents and propose the mechanisms controlling adsorption. The adsorption kinetics is examined using pseudo⁷³ and second-order⁷⁴ kinetic models given by:

Pseudo first order model:

$$\log(q_e - q_t) = \log q_e - \frac{K_1}{2.303} \cdot t \tag{9}$$

Pseudo second order model:

$$\frac{t}{q_t} = \frac{1}{K_2 \cdot q_e^2} + \frac{1}{q_e} \cdot t \quad (10)$$

Where q_t (mg/g) is the adsorbed amount at the time t (min), K_1 (min^{-1}) and K_2 (g/mg min) are the pseudo-first order and pseudo-second order rate constants respectively. The slope and intercept of the plots $\ln(q_e - q_t)$ vs time (t) are used to calculate the constants K_1 and q_e while K_2 and q_e are deduced from the plot of t/q_e vs. t (Figure 14). For the pseudo-first-order kinetic, the experimental data deviate from linearity, suggesting the inapplicability of the model for the present system. On the other hand, the pseudo-second order kinetic model presents the best correlation ($R = 0.999$), and the adsorption process is predominated by chemisorption. The kinetic constants K_1 , K_2 , and the calculated adsorption capacities deduced after modeling are grouped together in Table 7.

Conclusion

The removal of pollutants from wastewater is an environmental concern, particularly pharmaceutical effluents, which pose risks to human health and the ecosystem. In this regard, activated carbon constitutes a promising adsorbent for which optimization of adsorption is essential to maximize efficiency.

In this study, statistical methodology and Box–Behnken Response Surface Design were found to be effective and reliable in finding the optimal conditions for the adsorption of amoxicillin onto activated carbon. The response surface design with desirability function (df) was applied to evaluate the interactive effects of three independent variables namely pH, temperature and adsorbent dose on the response R (%).

The experimental data were treated by analysis of variance (ANOVA) and adjusted by a second order polynomial using multiple regression analysis. The optimal conditions: adsorbent dose (0.125 g), T (45°C) and pH (5.037) led to a maximum removal of amoxicillin of 89.99%. The isotherms were analyzed by different models and the Langmuir equation, gave the best fit to the equilibrium, where 142.85 mg/g of amoxicillin ($R^2 = 0.999$) at 25°C was removed from the solution. The pseudo-second order kinetic model gave the best correlation ($R^2 = 0.999$), and adsorption of amoxicillin is predominated by chemisorption.

This study demonstrated that activated carbon has a high adsorption capacity compared to adsorbents reported in the literature and the study of the photodegradation of Amoxicillin in the presence of a semiconductor constitutes a logical continuation of this study.

Authors Contribution

M.ABBAS conceived and designed the study, performed the experiments and analyzed the data. And M.Trari revise and correction of the manuscript. All authors reviewed and approved the final version of the manuscript.

Declaration of Conflicting Interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Funding

The author(s) received no financial support for the research, authorship, and/or publication of this article.

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Data Availability Statement

All data presented in this study are included in the manuscript by the corresponding author.

References

- Xu W, Gan Z, Li X, Zou S, Li P, Hu Z. Jun Li Occurrence and elimination of antibiotics at four sewage treatment plants in the Pearl River Delta (PRD). *South China Water Research*. 2007; 41(19):4526–4534.
- Abazari R, Mahjoub AR, Shariati J. Synthesis of a nano-structured pillar MOF with high adsorption capacity towards antibiotics pollutants from aqueous solution. *J Hazard Mater*. 2019;366:439–451.
- Qiu W, Sun J, Fang M et al. Occurrence of antibiotics in the main rivers of Shenzhen, China: association with antibiotic resistance genes and microbial community. *Sci Total Environ*. 2019;653: 334–341.
- Krzeminski P, Tomei MC, Karaolia P et al. Performance of secondary wastewater treatment methods for the removal of contaminants of emerging concern implicated in crop uptake and antibiotic resistance spread: review. *Sci Total Environ*. 2019; 648:1052–1081.
- Li Q, Jia R, Shao J, He Y. Photocatalytic degradation of amoxicillin via TiO₂ nanoparticle coupling with a novel submerged porous ceramic membrane reactor. *J Clean Prod*. 2019; 209:755–761.
- Roberto A, Marisa C, Raffaele M, Nicklas P. Antibiotic removal from wastewaters: the ozonation of amoxicillin. *J Hazard Mater*. 2005;122(3):243–250.
- Glen RB, Shaoyuan Z, Grimm DA. Naproxen removal from water by chlorination and *Biofilm Processes Water Res*. 2005, 39 (4): 668–676
- Vera H, Arminda A, Lúcia S. Microwave-assisted Fenton's oxidation of amoxicillin. *Chem Eng J*. 2013;220:35–44.
- Zümriye A, Özlem T. Application of biosorption for penicillin G removal: comparison with activated carbon. *Process Biochem*. 2005;40(2):831–847.
- Ettore Z, Sara C, Renzo B, Manuela M, Roberto F. Source, occurrence and fate of antibiotics in the Italian aquatic environment. *J Hazard Mater*. 2010:1042–1048.
- Elmolla ES, Chaudhuri M. Photocatalytic degradation of amoxicillin, ampicillin and cloxacillin antibiotics in aqueous

- solution using UV/TiO₂ and UV/H₂O₂/TiO₂ photocatalysis, *Desalination*. 2010;252(1–3):46-52
12. Eric KP, Ramon P, Jaka S, Nani I, Suryadi I. Performance of activated carbon and bentonite for adsorption of amoxicillin from wastewater: mechanisms, isotherms and kinetics. *Water Res*. 2009;43(9):2419-2430.
 13. Vera H, Arminda A, Lúcia S. Microwave-assisted Fenton's oxidation of amoxicillin. *Chem Eng J (Lausanne)*;220: 35-44.
 14. Amin K, Mahsa A, Mojtaba M et al. Monitoring of polycyclic aromatic hydrocarbons and probabilistic health risk assessment in yogurt and butter in Iran. *Food Sci Nutr*. 2021;9(4): 2114-2128.
 15. Michael I, Rizzo L, McArdell CS et al. Urban wastewater treatment plants as hotspots for the release of antibiotics in the environment. *A Review Water Resour*. 2013;47(3):957-995.
 16. Adriano J, Sebastian Z, Anke G et al. Biological degradation of pharmaceuticals in municipal wastewater treatment: proposing a classification scheme. *Water Res*. 2006;40(8): 1686-1696.ChristaS.
 17. Sourav KM, Amal KS, Alok S. Removal of ciprofloxacin using modified advanced oxidation processes: kinetics, pathways and process optimization. *J Clean Prod*. 2018; 171(10):1203-1214.
 18. Abdoallahzadeh H, Rashtbari Y, Américo-Pinheiro JHP, et al. Application of green and red local soils as a catalyst for catalytic ozonation of fulvic acid: experimental parameters and kinetic. *Biomass Conv. Bioref* 2023;1-10. <https://doi.org/10.1007/s13399-023-03895-6>
 19. José LS, Gabriel O, Araceli R, Silvia Á, José G, Juan G. Competitive adsorption studies of caffeine and diclofenac aqueous solutions by activated carbon. *Chem Eng J*. 2014;240: 443-453.
 20. Davood JN, Ali A, Nezam M et al. Parameters effecting on photocatalytic degradation of the phenol from aqueous solutions in the presence of ZnO nanocatalyst under irradiation of UV-C light. *Bulg Chem Commun*. 2015;47:14-18. Special Edition D.
 21. Gholamreza M, Ahamd A, Kamyar Y, Mahboube E. Preparation, characterization and adsorption potential of the NH₄Cl-induced activated carbon for the removal of amoxicillin antibiotic from water. *Chem Eng J*. 2013;217:119-128.
 22. Mohammad RS, Tariq JA-M, Anoushiravan M-B, Mansur Z. Adsorption of cephalixin from aqueous solution using natural zeolite and zeolite coated with manganese oxide nanoparticles. *J Mol Liq*. 2015;211:431-441.
 23. Sahmoune MN, Moussa A, Mohamed T. Understanding the rate-limiting step adsorption kinetics onto biomaterials for mechanism adsorption control. *Mechanism*. 2024;49:1-26.
 24. Fatemeh R, Maryam S, Shohreh A et al. The superior decomposition of 2,4-Dinitrophenol under ultrasound-assisted Fe₃O₄@TiO₂ magnetic nanocomposite: process modeling and optimization, Effect of various oxidants and Degradation pathway studies. *Int J Environ Anal Chem*. 2024;104(6):1243-1265.
 25. Ali A, Mohammad M, Kamyar Y et al. Magnetic NH₂-MIL-101(Al)/Chitosan nanocomposite as a novel adsorbent for the removal of azithromycin: modeling and process optimization. *Sci Rep*. 2022;12. Article 18990:1-16.
 26. Numchok M, Yuwalee U, Rameshprabu R. Bioethanol production from sunflower stalk: application of chemical and biological pretreatments by response surface methodology (RSM). *Biomass Conv Bioref*. 2021;11:1759-1773.
 27. John I, Pola J, Appusamy A. Optimization of ultrasonic assisted saccharification of sweet lime peel for bioethanol production using box-behnken method. *Waste Biomass Valor*. 2019;10: 441-453.
 28. Chan YT, Tan MC, Chin NL. Application of Box-Behnken design in optimization of ultrasound effect on apple pectin as sugar replacer. *LWT*. 2019;115:108449.
 29. Prola LDT, Machado FM, Bergmann CP et al. Adsorption of Direct Blue 53 dye from aqueous solutions by multi-walled carbon nanotubes and activated carbon. *J Environ Manag*. 2013; 130:166-175.AdebayoMA
 30. Glaydson Sdos R, Carlos HS, Eder CL, Michaela W. Preparation of novel adsorbents based on combinations of polysiloxanes and sewage sludge to remove pharmaceuticals from aqueous solutions. *Colloids Surf A Physicochem Eng Asp*. 2016 b;497: 304-331.
 31. Jagiello J, Thommes M. Comparison of DFT characterization methods based on N₂, Ar, CO₂, and H₂ adsorption applied to carbons with various pore size distributions. *Carbon*. 2004;42: 1227-1232.
 32. Thommes M, Kaneko K, Neimark AV, Olivier JP, Rodriguez-Reinoso F, Rouquerol J. Sing KSWPhysisorption of gases, with special reference to the evaluation of surface area and pore size distribution (IUPAC Technical Report). *Pure Appl Chem*. 2015; 87:1051-1069.
 33. Souza PR, Dotto GL. Salau NPG Artificial neural network (ANN) and adaptive neuro-fuzzy interference system (ANFIS) modeling for nickel adsorption onto agro-wastes and commercial activated carbon. *J Environ Chem Eng*. 2018;6(6): 7152-7160.
 34. Moussa A, Mohamed T. Contribution of zeolite to remove malachite green in aqueous solution by adsorption processes: kinetics, isotherms and thermodynamic studies. *Textil Res J*. 2023;93(15–16):3765-3776.
 35. Virkutyte J, Rokhina E, Jegatheesan V. Optimisation of Electro-Fenton denitrification of a model wastewater using a response surface methodology. *Bioresour Technol*.2010;101(5): 1440-1446
 36. Masoud Z, Wan Mo AWD, Mohamed KA. Optimization of the activity of CaO/Al₂O₃ catalyst for biodiesel production using response surface methodology. *Appl Catal, A*. 2009;366(1): 154-159.
 37. Ali A, Mohammad M, Kamyar Y et al. Magnetic NH₂-MIL-101(Al)/Chitosan nanocomposite as a novel adsorbent for the removal of azithromycin: modeling and process optimization. *Sci Rep*. 2022;12(18990):1-16.
 38. Gaddafi ID, Tawfik AS, Abdullahi AS. Response surface methodology optimization of adsorptive desulfurization on nickel/activated carbon. *Chem Eng J*. 2017;313:993-1003.

39. Jing H, Irene MCL, Guohua C. Comparative study of various magnetic nanoparticles for Cr(VI) removal. *Sep Purif Technol.* 2007;56(3):249-256.
40. Mohammad HM, Ali A, Anis J et al. Statistical modeling and optimization of dexamethasone adsorption from aqueous solution by Fe₃O₄@NH₂-MIL88B nanorods: isotherm, Kinetics, and Thermodynamic. *Environ Res.* 2023;236(Part 2):116773.
41. Langmuir I. The adsorption of gases on plane surface on glass, mica and platinum. *J Am Chem Soc.* 1918;40(9):1361-1403.
42. Freundlich H. Concerning adsorption in solutions. *Z Phys Chem Stoch.* 1906;57:385-470.
43. Temkin M, Pyzhev V, Kinetics of ammonia synthesis on promoted iron catalysts. *Acta Physicochim.* URSS 1940, 12: 327-356.
44. Hasan P, Hamid RG, Amir HM, Mina P, Ali A. Activated carbon derived from date stone as natural adsorbent for phenol removal from aqueous solution. *Desalination Water Treat.* 2017;72:406-417.
45. Valix M, Cheung WH, Zhang K, Role of heteroatoms in activated carbon for removal of hexavalent chromium from wastewaters. *J Hazard Mater* 2006, 135: 395-405.
46. Bruice PY. *Organic Chemistry.* International Edition; 2004.
47. Citraningrum M, Gunawan, Indraswati N, Ismadji S. Improved adsorption capacity of commercially available activated carbon norit row 0.8 Supra with thermal treatment for phenol removal. *J Environ Prot Sci.* 2007;1:62-74.
48. Kubilay S, Gurkan R, Savran A, Sahan T. Removal of Cu(II), Zn(II) and Co(II) ions from aqueous solutions by adsorption onto natural bentonite. *Adsorption.* 2007;13:41-51.
49. Foti C, Giuffrè O. Interaction of ampicillin and amoxicillin with Mn²⁺: a speciation study in aqueous solution. *Molecules.* 2020; 25(14):3110.
50. Proctor P, Gensmantel NP, Page MI. The chemical reactivity of penicillins and other β -lactam antibiotics. *J Chem Soc, Perkin Trans.* 1982;2(9):1185-1192.
51. Jean EH, Dominique M-L, Alejandro M et al. Factors essential for L,D-transpeptidase-mediated peptidoglycan cross-linking and β -lactam resistance in Escherichia coli. *Elife.* 2016;5:1-22.
52. Erah P, Goddard A, Barrett D, Shaw P, Spiller R. The stability of amoxicillin, clarithromycin and metronidazole in gastric juice: relevance to the treatment of Helicobacter pylori infection. *J Antimicrob Chemother.* 1997;39(1):5-12.
53. Dimitrakopoulou D, Rethemiotaki I, Frontistis Z, Xekoukoulotakis NP, Venieri D, Mantzavinos D. Degradation, mineralization and antibiotic inactivation of amoxicillin by UV-A/TiO₂ photocatalysis. *J Environ Manag.* 2012;98:168-174.
54. Elmolla ES, Chaudhuri M. Photocatalytic degradation of amoxicillin, ampicillin and cloxacillin antibiotics in aqueous solution using UV/TiO₂ and UV/H₂O₂/TiO₂ photocatalysis. *Desalination.* 2010;252(1-3):46-52.
55. Saucier C, Karthickeyan P, Ranjithkumar V, Lima EC, dos Reis GS, de Brum IAS. Efficient removal of amoxicillin and paracetamol from aqueous solutions using magnetic activated carbon. *Environ Sci Pollut Res.* 2017;24:5918-5932.
56. Thue PS, dos Reis GS, Lima EC et al. Activated carbon obtained from Sapelli wood sawdust by microwave heating for O-cresol adsorption. *Res Chem Intermed.* 2017a;43: 1063-1087.
57. Lavniewski A, de_Korwin JD, Muhale F, Jehl F. Gastric diffusion of antibiotics used against Helicobacter pylori. *Int J Antimicrob Agents.* 1998;9:181-193.
58. Dutta M, Dutta NN, Bhattacharya KG. Aqueous phase adsorption of certain beta-lactam antibiotics onto polymeric resins and activated carbon. *Sep Purif Technol.* 1999;16: 213-224.
59. Seyed YH, Ali A, Mohammad R, Kamyar Y. Application of Response Surface Methodology (RSM) in optimisation of fluoride removal by magnetic chitosan/graphene oxide composite: kinetics and isotherm study. *Int J Environ Anal Chem.* 2023;103(17):5368-5386.
60. Harrington Junior EC. The desirability function. *Ind Qual Control.* 1965;21:494.
61. Eric KP, Ramon P, Jaka S, Nani I, Suryadi I, Performance of activated carbon and bentonite for adsorption of amoxicillin from wastewater: mechanisms, isotherms and kinetics water research. 2009, 43: 2419-2430
62. Diana RL, Eder CL, Cibeles SU et al. Removal of amoxicillin from simulated hospital effluents by adsorption using activated carbons prepared from capsules of cashew of Para. *Environ Sci Pollut Control Ser.* 2019;26:16396-16408.
63. James MC, Philiswa NN. Effective adsorptive removal of amoxicillin from aqueous solutions and wastewater samples using zinc oxide coated carbon nanofiber composite. *Emerg Contam.* 2019;5:143-149.
64. Osvaldo P, André LC, Karen CB et al. NaOH-activated carbon of high surface area produced from guava seeds as a high-efficiency adsorbent for amoxicillin removal: kinetic, isotherm and thermodynamic studies. *Chem Eng J.* 2016;288:778-788.
65. Marcela AEde F, Cassandra Bde C, Mariana MB, Rafael de PS, Liliana AF. Removal of amoxicillin from water by adsorption onto activated carbon in batch process and fixed bed column: kinetics, isotherms, experimental design and breakthrough curves modeling. *J Clean Prod.* 2017;161: 947-956.
66. Marwa AC, Muthanna JA. Amoxicillin adsorption on microwave prepared activated carbon from *Arundo donax* Linn: isotherms, kinetics, and thermodynamics studies. *J Environ Chem Eng.* 2015;3(3):1592-1601.
67. Ali M, Maryam K, Hadi R, Roderick BW, Mahdi A. Amoxicillin removal from aqueous media using multi-walled carbon nanotubes. Fullerenes, nanotubes and carbon. *Nanostructures.* 2015;23(2):165-169.
68. Hamid F, Mehdi R, Mohammad AT, Ghasem S. Preparation of magnetic multi-walled carbon nanotubes for an efficient adsorption and spectrophotometric determination of amoxicillin. *J Ind Eng Chem.* 2015;21:889-892.
69. Adriana BS, Mariana RDS, Laura Mde A, Jose DA, Rochel ML, Maria HA. Iron oxide nanoparticles supported on mesoporous

- MCM-41 for efficient adsorption of hazardous β -lactamic antibiotics. *Water Air Soil Pollut.* 2018;229(59).
70. Jafari K, Heidari M, Rahmanian O. Corrigendum to "Wastewater treatment for Amoxicillin removal using magnetic adsorbent synthesized by ultrasound process. *Ultrason Sonochem.* 2018;45:248-256.
 71. Huaqing L, Zhen H, Hai L et al. Adsorption of amoxicillin by Mn-impregnated activated carbons: performance and mechanisms J. *RSC Adv.* 2016;6(14):11454-11460.
 72. Hu D, Wang L. Adsorption of amoxicillin onto quaternized cellulose from flax noil: kinetic, equilibrium and thermodynamic study. *J. Taiwan Inst Chem Eng.* 2016;64:227-234.
 73. Lagergren S. *Kung.* About the theory of so-called adsorption of soluble substances. *Sven Vetenskapsakad Handlingar.* 1898;24: 1-39.
 74. Ho YS, McKay G. Kinetic models for the sorption of dye from aqueous solution by wood. *J Environ Sci Health B.* 1998;76(4): 183-191.